Agents of pneumonia in slaughtered pigs in southern Brazil

Elisa R. De Conti, Karine L. Takeuti, Claiton I. Schwertz, Ronaldo M. Bianchi, David Driemeier, and David E.S.N. de Barcellos


A high prevalence of pneumonic lesions has been reported to affect slaughtered pigs in southern Brazil. In order to identify which microorganisms have been causing those lesions, 30 pig lungs presenting pneumonic gross lesions were collected from five different slaughterhouses, totaling 150 lungs. Samples for bacterial isolation, molecular, histopathologic and immunohistochemistry (IHC) evaluation were taken from each lung. The pneumonic lesion scoring ranged from 1.53 to 2.83. The most frequent histopathological lesions found was the concomitant Influenza A virus (IAV) and Mycoplasma hyopneumoniae infection, corresponding to 55.3% (83/150), and Pasteurella multocida type A was isolated in 54.2% (45/83) of these cases. In 102 samples (68%), there was histopathologic suggestion of involvement of more than one infectious agent. M. hyopneumoniae was the most frequent agent associated with pneumonic lesions, being present in 92.1% (94/102) of the lungs with coinfections, followed by IAV in 89.2% (91/102). Besides the coinfictions, IAV lesions were observed also in six samples without another pathogenic microorganism detected. A total of 46 samples with acute and subacute IAV suspected lesions in histopathological examination were assessed for IHC and real time RT-PCR for IAV. A total of 35% (16/46) of them were positive by IHC and 13% (6/46) by real time RT-PCR. Regarding M. hyopneumoniae, 79.3% (119/150) of samples were positive by qPCR and 84.9% (101/119) of them also presented M. hyopneumoniae suspected lesions in the histopathological examination. The results of this study suggest the importance of IAV in respiratory diseases in finishing pigs, even though this virus is more frequently reported in the nursery phase. In addition, our results emphasize the importance of lung coinfections in finishing pigs.

INDEX TERMS: Pneumonia, slaughter, pigs, Brazil, influenza A, Mycoplasma hyopneumoniae, Pasteurella multocida, coinfections, lungs, pneumonic lesions.
presente em 92,1% (94/102) dos pulmões com coinfecções, seguido de IAV, que foi encontrado em 89,2% (91/102). Além das coinfecções, lesões de IAV foram observadas em mais seis amostras que não apareciam envolvimento de outro agente infeccioso. Um total de 46 amostras com suspeita de lesão aguda e subaguda de IAV no exame histopatológico foram avaliadas para IHC e RT-PCR em tempo real para IAV e 35% (16/46) delas foram positivas por IHC e 13% (6/46) foram positivas por RT-PCR em tempo real. Com relação a M. hyopneumoniae, 79,3% (119/150) das amostras foram positivas por qPCR e 84,9% (101/119) delas também apresentaram lesões suspeitas de M. hyopneumoniae no exame histopatológico. Os resultados deste trabalho sugerem a importância do IAV como agente causador de pneumonias em suínos de terminação, embora esse vírus seja mais frequentemente relatado na fase de creche. Além disso, os achados deste trabalho demonstram a presença frequente de coinfecções pulmonares em suínos de terminação.

TERMS DE INDEXAÇÃO: Pneumonia, suínos, Brasil, influenza A, Mycoplasma hyopneumoniae, Pasteurella multocida, coinfecções, pulmões, lesões de pneumonia.

INTRODUCTION
Respiratory diseases correspond to a multifactorial problem in pigs and it is one of the major concerns to swine production. The economic losses are caused by increased pig mortality and intense use of antibiotics and vaccines and reduced daily feed gain and poor feed conversion rates (Haden et al. 2012). Pneumonic lesions are the most common finding in pigs lungs at slaughter in many countries (Fraile et al. 2010, Fablet et al. 2012, Brewster et al. 2017, Karabasil et al. 2017). The relationship between pneumonic lesions and losses of pig carcasses quality is also known, measured as a reduction of the post-trimming carcass weight in 1.26kg (Brewster et al. 2017).

The Porcine Respiratory Disease Complex (PRDC) is characterized by the coinfection of two or more bacterial or viral agents that act in synergism exacerbating the clinical signs especially in growing and finishing pigs. The occurrence of pneumonia at the end of the finishing phase in Brazilian pig herds is frequent. Studies of the last two decades showed a prevalence of pneumonic lesions in pigs at slaughter up to 75.7% (Silva et al. 2002). In a recent study evaluating the etiology of the lesion, especially pneumonic lesions in pigs at slaughter, as described by Markey et al. (2013). In the presence of pleuritis, necro-hemorrhagic areas of consolidation or nodules of necrosis in the lungs, microaerophilic incubation would be performed.

Lung lesion scoring. The severity index of pneumonic lesions was performed according to the methodology described by Madec & Kobisch (1982). Briefly, each pulmonary lobe contributes independently to the scoring, according to its proportion concerning the total lung area. The pulmonary lobes received a score from 0 to 4 according to the extent of the pulmonary consolidation lesion, where: 0 = normal lobe, without consolidation, 1 = from 1 to 25% of the lobe with consolidation, 2 = from 26% to 50%, 3 = from 51% to 75%, and 4 = from 76% to 100% of the affected lobe area with consolidation injury.

Bacterial isolation. Each sample was plated in sheep Blood Agar 5% (Kasvi®, Italy) and MacConkey Agar (Becton, Dickinson and Company®, USA) and incubated at 37°C for 24 to 48 hours in aerobic conditions. Biochemical tests were performed to identify the colonies, as described by Markey et al. (2013). In the presence of pleuritis, necro-hemorrhagic areas of consolidation or nodules of necrosis in the lungs, microaerophilic incubation would be performed.

Histopathological exam. After 24h of formalin fixation, each lung was routinely processed and embedded in paraffin blocks. Histological sections of 3-5µm were stained by hematoxylin-eosin (HE), as described by Allen (1992), for further description of the histopathologic lesions. All samples that were suspected of Influenza A virus were classified in an acute (suppurative necrotizing bronchiolitis/bronchitis), subacute (proliferative bronchiolitis/bronchitis associated with bronchointerstitial pneumonia), or chronic (bronchiolitis obliterans and fibrous tissue) lesion (Watanabe et al. 2012). Lung samples with BALT (bronchus-associated lymphoid tissue) hyperplasia were considered indicative of Mycoplasma hyopneumoniae infection (Blanchard et al. 1992). Suppurative bronchopneumonia with fibrin deposition was considered indicative of bacterial involvement in the etiology of the lesion, especially Pasteurella multocida.

Immunohistochemistry for Influenza A. Samples with acute and subacute lesions suggestive of Influenza virus infection in the histopathological evaluation were submitted to immunohistochemistry (IHC) for IAV. Briefly, the sample sections were placed on glass slides coated with poly-L-lysine and heated at 57°C until dry. The biotin-streptavidine-peroxidase Kit (KitsLab + System-HRP (Dako®, USA) was used to detect the presence of IAV. The endogenous activity of peroxidase was inhibited by immersion in the slides in methanol solution at 10% hydrogen peroxide for 10 minutes. Antigenic recovery was performed with enzymatic treatment using 0.05% protease XIV (Sigma®, USA) for 25 minutes at 37°C. All slides were incubated with primary antibodies Anti-IA (Milestone, dilution 1:800) overnight at 4°C. The chromogen was used as the 3,3-Diaminobenzidine (Dako®, USA) with Harris’ Hematoxylin.

MATERIALS AND METHODS

Ethics statement. The study was approved by the Institutional Animal Care and Use Committee of the “Universidade Federal do Rio Grande do Sul”, Brazil.

Study design and sampling. Five slaughterhouses under Federal Inspection Service (SIF), which slaughter pigs from five different companies (A, B, C, D, and E) located in the state of Rio Grande do Sul, Brazil, were selected for the study. Thirty lungs with macroscopic pneumonic lesions were randomly selected from two different batches in each slaughterhouse, totaling 150 lungs. The average age of the slaughtered animals was approximately 180 days. The lungs were collected for further score evaluation and sampling. Two fragments from transitional areas of each lung were sampled. One 4 cm x 4 cm fragment was collected and stored under refrigeration for bacteriological isolation and quantitative polymerase chain reaction (qPCR). A second fragment measured approximately 2cm3 was collected and fixed in a 10% neutral buffered formalin for histopathological and immunohistochemical examination.

Histopathological exam. After 24h of formalin fixation, each lung was routinely processed and embedded in paraffin blocks. Histological sections of 3-5µm were stained by hematoxylin-eosin (HE), as described by Allen (1992), for further description of the histopathologic lesions. All samples that were suspected of Influenza A virus were classified in an acute (suppurative necrotizing bronchiolitis/bronchitis), subacute (proliferative bronchiolitis/bronchitis associated with bronchointerstitial pneumonia), or chronic (bronchiolitis obliterans and fibrous tissue) lesion (Watanabe et al. 2012). Lung samples with BALT (bronchus-associated lymphoid tissue) hyperplasia were considered indicative of Mycoplasma hyopneumoniae infection (Blanchard et al. 1992). Suppurative bronchopneumonia with fibrin deposition was considered indicative of bacterial involvement in the etiology of the lesion, especially Pasteurella multocida.
as counterstain (Watanabe et al. 2012). The IAV positivity immune staining was assessed according to the structure, being classified as marked in the epithelium, bronchi and bronchioles, and macrophages.

**Real-time PCR for Influenza A.** Samples with acute and subacute lesions suggestive of Influenza infection in the histopathological evaluation were also submitted to real-time RT-PCR for IAV. Due to the short time for detection of the virus in lung tissue (Schaefer et al. 2013), samples with characteristic chronic lesions were not submitted to this test. The extraction of RNA was performed with a commercial extraction kit based on a silica column (AllPrep DNA/RNA Mini Kit, Qiagen®, Germany) as recommended by the manufacturer. The PCR reaction for IAV was performed from the extracted RNA, as described by the CDC (Centers for Disease Control, Atlanta, United States of America; World Health Organization) (WHO 2010). Samples were evaluated in duplicate, and two positive and two negative controls were added to each analysis.

**Real-time PCR for Mycoplasma hyopneumoniae.** The DNA extraction was performed with a commercial extraction kit based on a silica column (Purelink Genomic DNA Mini kit, Thermo Fisher Scientific®, USA) following manufacturer’s instructions. The PCR was performed in all samples, according to Takeuti et al. (2017). The samples were evaluated in duplicate, and two additional positive and negative were added to each analysis.

**Statistical analysis.** Multiple pairwise comparison tests of proportion with Bonferroni correction were performed in R v3.2 (R Core Team 2013) to compare the prevalence of positive lungs for *P. multocida* by isolation and the prevalence of positive samples for IAV and *M. hyopneumoniae* by PCR among the companies. Chi-square analysis was performed to compare absence or presence of histopathological lesions, bacterial isolation and PCR results. Statistical significance was considered when *P*-values were less than 0.05.

### RESULTS

**Pulmonary scoring at slaughter.** All selected lungs presented macroscopic pneumatic lesions. The only pattern of lesion observed was cranioventral consolidation (Fig.1A).

The mean lung lesion score ranged from 1.53 to 2.83 among companies (Table 1). No lungs (0/150) had pleuritis, necrohemorrhagic areas of consolidation or nodules of necrosis.

**Bacteriology.** *Pasteurella multocida* type A was the only bacterium isolated with relevance to pneumatic lesions. The bacterium was isolated in 43.3% (65/150) of the lungs, and company C did not have any lung sample with *P. multocida* type A isolation. However, companies D and E presented a higher number of lungs with *P. multocida* type A isolation when compared to companies A and C (Table 1).

**Histopathological evaluation.** There was a predominance of lung lesions suggestive of *Mycoplasma hyopneumoniae* and Influenza A virus (IAV) coinfection, representing 55.3% (83/150) of the lungs, followed by 28% (42/150) of samples suggestive of single *M. hyopneumoniae* infection (Fig.1B), and 9.3% (14/150) of samples indicative of single IAV infection.

### Table 1. Lung lesion scoring and distribution of positive lungs for *Pasteurella multocida* type A isolation, and *Mycoplasma hyopneumoniae* and Influenza A virus suggestive lesions by histopathological evaluation in five companies assessed in the study

<table>
<thead>
<tr>
<th>Company</th>
<th>A</th>
<th>B</th>
<th>C</th>
<th>D</th>
<th>E</th>
</tr>
</thead>
<tbody>
<tr>
<td>IAV</td>
<td>86.7%*</td>
<td>70.0%*</td>
<td>20.0%*</td>
<td>76.7%*</td>
<td>70.0%*</td>
</tr>
<tr>
<td>(26/30)</td>
<td>(21/30)</td>
<td>(6/30)</td>
<td>(23/30)</td>
<td>(21/30)</td>
<td></td>
</tr>
<tr>
<td>Mhyo</td>
<td>80.0%**</td>
<td>86.6%**</td>
<td>73.3%*</td>
<td>90.0%*</td>
<td>86.6%**</td>
</tr>
<tr>
<td>(24/30)</td>
<td>(26/30)</td>
<td>(22/30)</td>
<td>(27/30)</td>
<td>(26/30)</td>
<td></td>
</tr>
<tr>
<td>PmA</td>
<td>33.3%*</td>
<td>50.0%*</td>
<td>0%*</td>
<td>66.7%*</td>
<td>66.7%*</td>
</tr>
<tr>
<td>(10/30)</td>
<td>(15/30)</td>
<td>(0/30)</td>
<td>(20/30)</td>
<td>(20/30)</td>
<td></td>
</tr>
<tr>
<td>LLS</td>
<td>2.03</td>
<td>2.40</td>
<td>1.53</td>
<td>2.83</td>
<td>2.23</td>
</tr>
</tbody>
</table>

IAV = Influenza A virus, Mhyo = Mycoplasma hyopneumoniae, PmA = Pasteurella multocida type A, LLS = lung lesion score; Different superscript letters within a row indicate statistically significant difference (*P*<0.05) among the five companies (A, B, C, D, and E) based on multiple pairwise comparison tests of proportion analysis.

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![Fig1.](image)

**Fig.1.** (A) Lung of a pig selected for the study, dorsal view. Multifocal to coalescent, reddish and firm areas in the cranial and middle lobes are observed (consolidation). The cuts in the diaphragmatic lobe were performed by the slaughterhouse inspection. (B) Histological aspect of the lung of a pig selected for the study. There is bronchointerstitial pneumonia associated with BALT (bronchus-associated lymphoid tissue) hyperplasia, edema and mild inflammatory infiltrate of macrophages in alveolar spaces. These lesions were observed in cases of *Mycoplasma hyopneumoniae* pneumonia. HE, obj.20x. (C) Histological aspect of the lung of a pig selected for the study. An area of bronchiolitis obliterans is observed associated with moderate inflammatory infiltrate of neutrophils and macrophages in alveolar and bronchiolar lumen. There is also type II pneumocytes hyperplasia, and interstitial inflammatory infiltrate of lymphocytes and plasma cells. These lesions were considered compatible with chronic Influenza A pneumonia. HE, obj.40x.
(Fig. 1C). Moreover, 7.4% (11/150) of lung samples did not show any suspected lesion in the histopathological examination. Taking all histopathological results into account, a total of 64.7% (97/150) of them resulted in microscopic lesions suggestive of IAV infection, isolated or in a coinfection with *M. hyopneumoniae*. The association of histological lesions and *P. multocida* type A isolation is summarized in Table 2.

**Immunohistochemistry and real-time RT-PCR for Influenza A**. A total of 46 samples with acute and subacute IAV suspected lesions in histopathological examination were selected for IHC and real time RT-PCR for IAV. A proportion of 35% (16/46) of them were positive by IHC with macrophage marking, and 13% of the samples (6/46) were positive by real time RT-PCR.

**Detection of *M. hyopneumoniae* by real-time PCR (qPCR)**. A total of 79.3% (119/150) of samples were positive for *M. hyopneumoniae* by qPCR. Companies A, B, D and E had similar results, ranging from 90% (A) to 100% (D) of positives samples, and the majority of samples also presented histopathological lesions suggestive of *M. hyopneumoniae* infection. On the other hand, company C presented only 16.6% (5/30) of positives samples by qPCR (Table 3).

**DISCUSSION**

All selected lungs of this study showed gross pneumonic lesions, as expected since it was a selection criterion in each slaughterhouse. The lungs of all companies showed high lesion scores, but company C presented the lowest scoring, but company C presented the lowest scoring, and company D presented the highest scoring, as expected since it was a selection criterion in each company.

A total of 79.3% (119/150) of samples which presented *P. multocida* type A isolation also showed histopathological lesions suggestive of *M. hyopneumoniae* infection. On the other hand, company C presented only 16.6% (5/30) of positives samples by qPCR (Table 3).

**Table 2. Relationship between *Pasteurella multocida* type A isolation and histopathological lesions suggestive of IAV and *Mycoplasma hyopneumoniae* coinfections**

<table>
<thead>
<tr>
<th>Histological aspect</th>
<th>Companies</th>
<th>Mhyo qPCR</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>Bronchointerstitial pneumonia with BALT hyperplasia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>70% (21/30)</td>
<td>10% (3/30)</td>
</tr>
<tr>
<td>B</td>
<td>80% (24/30)</td>
<td>6.7% (2/30)</td>
</tr>
<tr>
<td>C</td>
<td>13.3% (4/30)</td>
<td>60% (18/30)</td>
</tr>
<tr>
<td>D</td>
<td>90% (27/30)</td>
<td>0% (0/30)</td>
</tr>
<tr>
<td>E</td>
<td>83.3% (25/30)</td>
<td>3.3% (1/30)</td>
</tr>
<tr>
<td>Total</td>
<td>67.3% (101/150)</td>
<td>16% (24/150)</td>
</tr>
</tbody>
</table>

**Table 3. An overall view of correspondence of lungs with histopathologic *Mycoplasma hyopneumoniae* (Mhyo) suspected lesions and *M. hyopneumoniae* qPCR results regarding the five companies enrolled in the study**

<table>
<thead>
<tr>
<th>Histological aspect</th>
<th>Companies</th>
<th>Mhyo qPCR</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>No evidence of BALT hyperplasia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>20% (6/30)</td>
<td>0% (0/30)</td>
</tr>
<tr>
<td>B</td>
<td>13.3% (4/30)</td>
<td>0% (0/30)</td>
</tr>
<tr>
<td>C</td>
<td>3.3% (1/30)</td>
<td>23.3% (7/30)</td>
</tr>
<tr>
<td>D</td>
<td>10% (3/30)</td>
<td>0% (0/30)</td>
</tr>
<tr>
<td>E</td>
<td>13.3% (4/30)</td>
<td>0% (0/30)</td>
</tr>
<tr>
<td>Total</td>
<td>12% (18/150)</td>
<td>4.7% (7/150)</td>
</tr>
</tbody>
</table>

qPCR = quantitative polymerase chain reaction, BALT = bronchus-associated lymphoid tissue; Different letters within a row indicate a statistically significant difference (P<0.05) based on chi-square analysis.
proportion of lungs positive for *M. hyopneumoniae* by PCR. This data emphasizes the importance of this bacterium in Brazilian finishing pigs, as previously reported (Takeuti et al. 2013, Vicente et al. 2013, Morés et al. 2015), which could be related to the high density of pig farms in Brazilian pig production areas and the very high prevalence of positive farms for *M. hyopneumoniae* in the country. It is important to note that *P. multocida* type A isolation was significantly lower in samples from company C when compared to all other companies. This may be related to the lower number of positive lungs for *M. hyopneumoniae* by qPCR.

Regarding histopathological analysis, 83.3% of samples presented BALT (bronchus-associated lymphoid tissue) hyperplasia, although not all of them were positive on qPCR for *M. hyopneumoniae*. The high concordance of 67.3% (101/150) between qPCR positivity and histologic lesions suggestive of *M. hyopneumoniae* was expected, once BALT hyperplasia is a characteristic finding of fully developed mycoplasma pneumonia (Blanchard et al. 1992, Caswell & Williams 2016). However, in 16% (24/150) of the samples there was a disagreement between the presence of BALT hyperplasia and the lack of *M. hyopneumoniae* qPCR detection which could be due to the fact that BALT hyperplasia is not a pathognomonic histopathologic lesion for *M. hyopneumoniae*, and can be seen in cases of pneumatic lesions caused by other agents as IAV (Laisse et al. 2018). In addition, a total of 12% (18/150) of the samples were *M. hyopneumoniae* positive by qPCR without presenting BALT hyperplasia. As *M. hyopneumoniae* has a very chronic and long-term infection in pigs (Pieters & Maes 2019), the detection of these microorganism without histopathologic lesions could be associated with a recent or very mild infection with not enough time to cause microscopic lesions, such as BALT hyperplasia. Additionally, in this study, bronchointerstitial pneumonia was a very common finding and could be attributed both to *M. hyopneumoniae* and IAV (Schaefer et al. 2013, Caswell & Williams 2016).

After the emergence of pandemic H1N1 Influenza A virus (IAV) in 2009, the IAV has been reported as an important agent of the PRDC in Brazilian swine production (Rajão et al. 2013, Rech et al. 2018). The presence of pigs showing clinical signs of IAV infection is frequently associated with the nursery phase, probably due to the decrease of the levels of maternal immunity and the increase of infection susceptibility (Van Reeth & Vincent 2019). Ciacci-Zanella et al. (2015) found an average prevalence of antibodies against IAV in Brazilian pig herds of 78.1% in the nursery phase. However, in our work we found a high prevalence of IAV suggestive histopathological lesions (64.6%) also in finishing pigs. All companies showed high frequency of positive samples, which ranged from 70% to 86.7%, except for company C, which presented the lowest result (20%). As the IAV infection has a fast resolution, with convalescence around seven days after infection (Schaefer et al. 2013), and the histopathologic lesions resolution occurs within three weeks after infection (Janke 2000), the IAV lesions found in this work probably occurred during the finishing phase, which is not commonly reported. The gross lesions found in uncomplicated IAV infection are limited to the apical and cardiac lobes of the lung (Van Reeth & Vincent 2019), the same pulmonary lobes affected by *M. hyopneumoniae*, which may result in misleading diagnosis when laboratory tests are not performed. The similarity of subacute or chronic histopathologic lesions of IAV with initial lesions of *M. hyopneumoniae* infection, especially in cases where the lymphocytic infiltrate forms a ring around the bronchioles (Janke 2014), may confuse the diagnostics between these microorganisms by histopathologic examination. Taking this fact into account, we submitted samples with IAV subacute and acute histopathological lesions to IHC and real-time RT-PCR for IAV and we observed a total of 35% and 13% of positive results for IAV, respectively. Detection of IAV has been reported to be possible for no more than 11 days post infection (p.i.) by real-time RT-PCR (Lange et al. 2009) and 13 days p.i. by IHC (Weingartl et al. 2010). This difference could justify the higher detection by IHC than real time RT-PCR in our study. Another explanation could be related to the multifocal distribution of lesions or the condition of the samples (Vincent et al. 1997), since they were collected from different areas of the lungs. These findings emphasize the importance of laboratory diagnostic of pig pneumonia and the relevance of IAV infection in pigs also during the finishing phase, which should be considered in the differential diagnosis of pneumatic lesions in pigs. In addition, more studies regarding the infection dynamics of this virus in Brazilian pig herds, especially during the finishing phase should be performed.

Although other microorganisms have been reported to contribute to the Porcine Respiratory Disease Complex (PRDC), only IAV, *M. hyopneumoniae* and *P. multocida* type A were found in this work. The lack of additional histopathological lesions suggestive of other agents was the reason for no further tests. No evidences of PCV2 infection were observed in all lungs assessed in this study, which was expected since the virus usually affects younger pigs, and due to the good control of infection by commercial vaccines usage (Morés et al. 2015). In addition, PRRSv and PRCV are important virus in the world pig industry but they have never been reported in Brazil (Rech et al. 2018), so the absence of suggestive lesions for the agents was expected.

The presence of more than one infectious agent within all lungs from five companies was detected in 68% of the samples regarding histopathological, bacteriological and molecular findings. This data demonstrated the importance of coinfections in pig lungs at slaughter age, being *M. hyopneumoniae*, IAV and *P. multocida* type A the three most frequent microorganisms identified in our study. Although *M. hyopneumoniae*, *P. multocida* and Influenza A virus had already been describe as the most relevant aggravating of PRDC in Brazil within pigs from all ages (Rech et al. 2018), the high frequency of IAV in slaughtered pigs was an unexpected finding, as previous work have observed only a 2.8% frequency in fattening pigs (Morés et al. 2015). These findings have an important economic impact, since the economic losses due to coinfections between IAV and *M. hyopneumoniae* were estimated to be 16 times greater than single infections caused only by *M. hyopneumoniae* (Haden et al. 2012). Furthermore, our results suggest that those microorganisms should be taken into account in order to control pig pneumonia at the end of the finishing phase in Brazilian pig farms.

**CONCLUSION**

The data of this work highlight the coinfection roll of *Mycoplasma hyopneumoniae*, Influenza A virus (IAV) and *Pasteurella multocida* type A as the main agents of the Porcine Pneumonia.
Respiratory Disease Complex in pigs from Brazil. Moreover, we highlight the high frequency of lesions caused by IAV and the identification of the virus by real time RT-PCR in pigs at slaughter age, even though IAV is not typically associated with infections and clinical cases in pigs in this phase.

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Conflict of interest statement.- The authors declare that they have no competing interest.

REFERENCES


