



Histological characterization of reproductive tract and fetal annexes of the West Indian Manatee (*Trichechus manatus*) from Brazil¹

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ABSTRACT.- Bezerra A.R., Salmito-Vanderley C.S.B., Bersano P.R.O., Carvalho V.L., Meirelles A.C.O., Attademo F.L.N., Luna F.O. & Silva L.D.M. 2018. **Histological characterization of reproductive tract and fetal annexes of the West Indian Manatee (*Trichechus manatus*) from Brazil.** *Pesquisa Veterinária Brasileira* 38(11):2166-2174. Universidade Estadual do Ceará, Avenida Dr. Silas Munguba 1700, Campus do Itaperi, Fortaleza, CE 60714-903, Brazil. E-mail: lucia.daniel.machado@hotmail.com

The West Indian manatee (*Trichechus manatus*) is one of the most threatened aquatic mammals in Brazil, and is currently classified as “endangered” (MMA). The objective of this study was to characterize histologically the reproductive tract and fetal annexes of stranded manatees in northeastern Brazil. Tissue samples were collected from the reproductive tract of 23 manatees, which were fixed in 10% buffered formalin, processed using standard histological protocols and stained with hematoxylin eosin. We qualitatively described the histological and histomorphometric characteristics of each structure. Six ovaries were analyzed. In four ovaries, we found a large number of primordial and primary follicles. Two ovaries were different from the others: one had inflammatory infiltration and the other had a thickening in the cortex and absence of follicles. We also analyzed seven uteri (of which four were in the proliferative phase, two in the secretory phase, and one in the recovery phase), four placentas, one vagina, six testes (four were in the immature phase, one in the pubertal phase, and one in the mature phase), two epididymides, two penises, and one umbilical cord. The histological and morphometric findings in our work will support future analyses of the reproductive tract of *T. manatus* from Brazil.

INDEX TERMS: Histology, reproductive tract, fetal annexes, West Indian Manatee, *Trichechus manatus*, Brazil, sirenian, reproductive biology, embryonic annexes.

RESUMO.- [Caracterização histológica do trato reprodutor e anexos fetais do peixe-boi marinho (*Trichechus manatus*) do Brasil.] O peixe-boi marinho (*Trichechus manatus*) é um dos mamíferos aquáticos mais ameaçados do Brasil e, atualmente é classificado como “em perigo” (MMA).

O objetivo deste estudo foi caracterizar histologicamente o trato reprodutor e os anexos fetais de peixes-bois marinhos encalhados no Nordeste do Brasil. Foram coletadas amostras de tecidos do trato reprodutor de 23 peixes-bois marinhos (*T. manatus*), que foram fixadas em formol tamponado a 10%, processados usando protocolos histológicos padrão e corados com hematoxilina eosina. Foi realizada a descrição qualitativa das características histológicas e histomorfométricas de cada estrutura. Foram analisados seis ovários. Em quatro ovários, foi encontrado um grande número de folículos primordiais e primários. Dois ovários eram diferentes dos outros: um tinha infiltração inflamatória e o outro tinha um espessamento no córtex e ausência de folículos. Também foram analisadas sete amostras uterinas (das quais quatro estavam na fase proliferativa, duas na fase secretória e uma

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na fase de recuperação), quatro placentas, uma vagina, seis testículos (quatro na fase imatura, um na fase puberal e um na fase madura), dois epidídimos, dois pênis e um cordão umbilical. Os achados histológicos e morfométricos em nosso trabalho apoiarão futuras análises do trato reprodutivo de *T. manatus* do Brasil.

TERMOS DE INDEXAÇÃO: Histologia, trato reprodutor, anexos fetais, peixe-boi marinho, *Trichechus manatus*, Brasil, sirênio, biologia reprodutiva, anexos embrionários.

INTRODUCTION

The population of manatees (*Trichechus manatus*) in the Brazilian Northeast has been estimated to range from 485 to 2,221 individuals, based on overflight measurements, which indicates that there are few manatees in this region (Alves et al. 2016). This species represents one of the most endangered aquatic mammals in Brazil, and is classified as “endangered” in the Official National List of Endangered Species (Brasil 2014).

Some issues faced by the manatee population of the Brazilian Northeast include the rapid growth of anthropogenic activities in the coastal zones, habitat destruction owing to the loss of mangroves, estuarine sedimentation (leading females to have their offspring in more agitated waters, risking the cubs to be carried away by tides and stranded on beaches), water pollution, and debris ingestion; because manatees have coastal habits and this area has a higher concentration of litter (Parente et al. 2004, Meirelles 2008, Lima et al. 2011, Attademo et al. 2015).

To develop effective conservation strategies for endangered species, such as the manatee, it is crucial to understand their life history and reproductive biology (Marsh et al. 2011). Studies on manatees from the United States, where a large number of dead animals were evaluated, resulted in an analysis of the reproductive tracts of males and females (Hernandez et al. 1995, Marmontel 1995). According to these authors, male manatees present a vascular penis that retracts when not engorged, do not have a crotch, have intra-abdominal testicles (with a ventrolateral position to the kidneys), and bilateral seminal vesicles, which are in a dorsal position in relation to the urinary bladder. However, the prostate is composed of erectile muscle tissue (Reynolds III et al. 2002).

Regarding the females, their reproductive system is composed of ovaries, uterine tubes, uterus, vagina, vaginal vestibule, and clitoris. The mammary glands also contribute to the reproductive process. Female manatees have a bicornuate uterus, and with a zonary (anatomical classification) and endotheliochorial (histological classification) placenta (Rodrigues et al. 2008).

Although this species has been studied for more than three decades, research on its reproductive system has not yet been performed in Brazil. The histological evaluation of the reproductive system has been carried out by the analysis of samples collected from stranded dead animals. However, access to these types of samples has been limited in Brazil because as the small size of the population, the high frequency of newborn stranding, the stage of decomposition

of dead stranded individuals and the storage of samples in the collections (Meirelles & Carvalho 2016).

Thus, the objective of this study was to characterize the reproductive tract and fetal annexes of male and female manatees stranded in northeastern Brazil to support future studies on the age of sexual maturation in these animals.

MATERIALS AND METHODS

Ethical aspects. This experiment was approved by the Ethics Committee for Animal Use of the State University of Ceará (number 0971110/2016) and the Biodiversity Authorization and Information System (SISBIO)/Chico Mendes Institute for Biodiversity Conservation (ICMBio, license number 51456-3).

Animals. We analyzed tissue samples from the reproductive tracts of 23 manatees (*Trichechus manatus* Linnaeus, 1758), including 14 females and 9 males of different age groups. These animals were stranded, died in rehabilitation, or gave birth in captivity from July 1995 to October 2016 (Tables 1 and 2). Manatees were necropsied according to conventional protocols (Geraci et al. 2005). Samples were included in the biological collections of the Association for Research and Preservation of Aquatic Ecosystems (Aquisis) in Caucaia, Ceará and the National Center for Research and Conservation of Aquatic Mammals (CMA/ICMBio) in Itamaracá, Pernambuco.

Histological processing. Tissues from the reproductive tracts were fixed in 10% buffered formalin. We collected tissue samples approximately 2cm-thick and processed them using standard histological protocol before histological staining with hematoxylin eosin (HE). Slides containing the processed samples were analyzed using light microscopy. During the histological processing, samples were dehydrated through successive washing in alcohol solutions with gradually increased concentrations (70, 80, 95, and 100%). Samples were washed for 50 min with each solution and then maintained in an alcohol/xylol solution for 24h. Thereafter, samples were diaphanized, where they were successively placed in two xylol solutions for 50 min each. They were then subjected to an inclusion process, in which samples were impregnated with two sets of paraffin for 50 min at 60°C. We then embedded the biological material in histological paraffin.

The embedded samples were cut into 5µm-thick sections using a histological resin microtome (Leica RM 2235). Sections were then immersed in a histological water bath at a mean temperature of 45°C and collected on slides pre-prepared with albumin. The slides containing the sections were maintained in an oven (MD 1.2) for sterilization and drying. Next, the slides were histologically stained with hematoxylin eosin. Fragments of testis, penis, umbilical cord, and placenta tissue were later stained with Masson's trichrome.

After this procedure, slides were analyzed using light microscopy and were photomicrographed (Nikon Eclipse 80i) using a digital photomicrographic system (Nikon NIS-Elements Advanced Research). During the analysis, we qualitatively described the histological and histomorphometric characteristics of each structure: a) ovaries (cortex thickness, largest and smallest diameter of the follicles with an observable oocyte); b) uterus (thickness of mucosa and endometrium); c) vagina (thickness of the vaginal epithelium), and d) testicles (major and minor diameter of the seminiferous tubules). The analysis was performed using ImageJ software, with an average of ten measurements for each structure.

Statistical analysis. Subjective data were presented in descriptions, whereas objective data were expressed as mean ± standard deviation.

Table 1. Information on the female manatees (*Trichechus manatus*) analyzed

Register number	Date of stranding or death	Place of stranding	Total length (meters)	Age group	Samples
02S0112/31	14/11/2005	Fontainhas, Aracati/CE	3.15	Adult	Uterus
02S0112/36	26/09/2006	Between Redonda and Peroba, Icapuí/CE	3.07	Adult	Ovary
02S0112/38	25/02/2007	Retiro Grande, Icapuí/CE	3.18	Adult	Uterus
02S0112/57	19/10/2012	Canto Verde, Beberibe/CE	1.20	Calf	Uterus
02S0112/60	21/09/2013	Retirinho, Aracati/CE	2.41	Adult	Ovary
02S0112/77	17/10/2016	Ponta Grossa, Icapuí/CE	3.18	Adult	Ovary
08S0112/20	24/03/2015	Alagamar, Grossos/RN	1.20	Calf	Ovary and uterus
01S0112/02	21/01/2008	Barro Preto/CE	3.07	Adult	Placenta
01S0112/07	08/06/2007	Ponta de Pedra/PE	2.90	Adult	Placenta
01S0112/08	30/07/2008	Barra de Sucatinga, Beberibe/CE	2.89	Adult	Placenta
01S0112/12	28/07/1995	Jacarecica, Maceió/AL	1.08	Calf	Ovary, uterus and vagina
01S0112/25	11/04/2008	Born in captivity, 10/4/97, PE	2.59	Adult	Placenta
01S0112/30	10/11/1999	Barra de Camarutuba/PB	1.36	Calf	Uterus
01S0112/31	02/03/1998	Pitimbi/PB	-	-	Ovary and uterus

The age group was defined according to Marmontel (1995).

Table 2. Information on the male manatees (*Trichechus manatus*) analyzed

Register number	Date of stranding or death	Place of stranding	Total length (meters)	Age group	Samples
02S0111/42	13/02/2008	Redonda, Icapuí/CE	1.19	Calf	Testicles
02S0111/45	10/10/2008	Parajuru, Beberibe/CE	1.15	Calf	Testicles
02S0111/55	23/09/2011	Ponta Grossa, Icapuí/CE	2.01	Juvenile	Testicles
02S0111/59	23/03/2013	Manibu, Icapuí/CE	1.08	Calf	Testicles and epididymis
02S0111/71	05/03/2015	Ariós, Beberibe/CE	1.35	Calf	Epididymis
02S0111/73	03/10/2015	Ponta Grossa, Icapuí/CE	3.01	Adult	Testicles and penis
01S0111/123	04/11/2002	Born in captivity/PE	1.06	Calf	Testicles
01S0111/180	24/12/2004	Barretas, Nisia Floresta/RN	2.72	Adult	Penis
01S0111/304	16/11/2011	Born in captivity/PE	0.79	Calf	Umbilical cord

The age group was defined according to Marmontel (1995).

RESULTS

We analyzed six ovaries, two of which were different from the others. We also analyzed seven uteri, four placentas, one vagina, six testicles, two epididymides, two penises, and one umbilical cord. These tissues were collected from manatees of different sizes and age groups (Table 1 and 2).

Ovaries

The ovary was coated with a serum layer consisting mainly of simple cubic epithelium, which, in some cases, had features of pseudostratified epithelium or small areas of columnar epithelium (Fig.1A).

The ovarian cortex consisted of a narrow band (Table 3) with the presence of follicles surrounded by supporting tissue. In manatees 08S0112/20, 01S0112/12, and 01S0112/31, there was a predominance of primordial follicles, but some primary follicles were also observed (Fig.1B). A secondary and a tertiary follicle were also observed in manatee 01S0112/31. In the ovary fragment of manatee 02S0112/60, we detected primordial, primary, secondary, tertiary, and Graafian follicles (Fig.1C).

This organ contained a large area of bone marrow (approximately 80% of the ovary), which consisted of loose connective tissue with many adipocytes and was richly and diffusely vascularized with vessels of different calibers (Fig.1D).

At the time of the necropsy of manatee 02S0112/36, the ovaries were not found. However, we observed the presence of tissue in the left antimer, with discrete signals indicative of possible ovarian tissue. We did not find similar tissue or ovarian tissue in the right antimer. In the ovary of manatee 02S0112/36, the cortex presented a thicker region (403,882µm) than in the ovaries of the other analyzed manatees, containing almost no follicle. In addition, this ovary presented considerable connective tissue in the supporting tissue of the cortex, where we observed a band with an intense increase in the cellularity composed of dense connective tissue and another band with less cellularity composed of loose connective tissue (Fig.1E). The ovarian marrow in manatee 02S0112/36 was similar to that observed in the ovaries of the other analyzed manatees.

In the ovary of manatee 02S0112/77 (supposedly an adult female), we found a large number of blood vessels, histiocytic inflammatory infiltrate (predominance of macrophages),

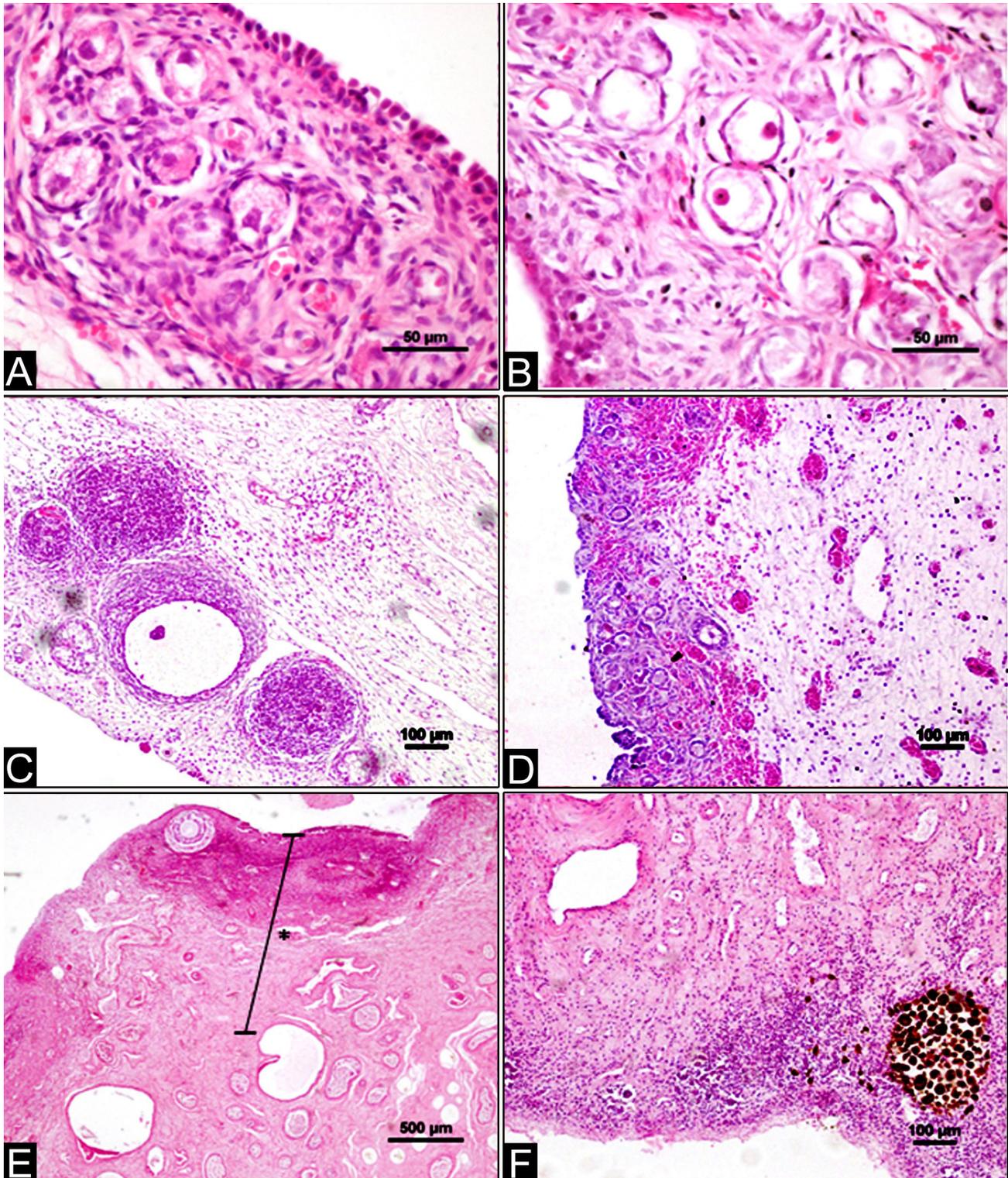


Fig.1. Ovary of *Trichechus manatus*. (A) Narrow cortex and germinal epithelium. HE, obj.40x. (B) Primary follicles. HE, obj.40x. (C) Tertiary follicle. HE, obj.10x. (D) Narrow cortex and large marrow area, highly vascularized. HE, obj.10x. (E) Altered ovary of manatee 02S0112/36 with thickened cortex. HE, obj.40x. (F) Ovary of manatee 02S0112/77 with an inflammatory infiltrate. HE, obj.10x.

multifocal necrosis, coalescent and congested vessels, congestion areas in the periphery, and the presence of some follicles (primary and secondary), which were in an advanced degeneration state (Fig.1F).

Uterus

The uterine mucosa was lined with simple cylindrical epithelium followed by endometrial glands with the same coating, in a perpendicular position to the mucosa. The submucosa

Table 3. Mean \pm standard deviation (SD) of the epithelium, cortex, primordial follicle major diameter (PFMa), primordial follicle minor diameter (PFMi), primary follicle major diameter (PaFMa), primary follicle minor diameter (PaFMi), secondary follicle major diameter (SFMa), and secondary follicle minor diameter (SFMi) measured in μm in female manatees (*Trichechus manatus*)

Ovary	Epithelium	Cortex	PFMa	PFMi	PaFMa	PaFMi	SFMa	SFMi
01S0112/12	18.262	192.438	43.149	38.162	42.127	39.927	ND	ND
08S0112/20	12.592	197.413	42.583	34.218	51.648	45.825	ND	ND
01S0112/31	9.983	200.277	43.274	35.250	55.656	41.880	ND	ND
02S0112/60	12.346	291.739	45.260	37.087	51.195	47.580	124.960	95.270
02S0112/36	16.364	403.882	ND	ND	ND	ND	ND	ND
02S0112/77	10.544	200.050	ND	ND	ND	ND	ND	ND
Mean \pm SD	13.348 \pm 3.287	247.633 \pm 85.363	43.567 \pm 1.126	36.179 \pm 1.776	50.156 \pm 5.716	43.803 \pm 3.516	124.96 \pm 0	95.27 \pm 0

ND = no data.

was composed of abundant fibrocellular and fibrovascular tissue (Table 4). The muscular layer was composed of three layers, longitudinal, circular, and longitudinal. The serosa was composed of mesothelium-lined connective tissue. The uteri of the manatees 08S0112/20, 01S0112/12, 01S0112/31, and 02S0112/31 were in the proliferative phase (Fig.2A). The uterus of the manatees 01S0112/30 and 02S0112/57 were in the secretory phase (Fig.2B), with glands opening into the lumen of the organ. The uterus of the manatee 02S0112/38 was in the recovery phase (Fig.2C).

Placenta

In the placenta, we detected the presence of an artery in the fetal chorion, and a vein. We also observed a well-developed villus region (lacunar region with the presence of a syncytial region, syncytiotrophoblast), and the presence of maternal tissue cells with chorionic villi (conjunctival and mesoderm). We also observed connective tissue filled with blood and maternal tissue (Fig.2D).

Vagina

The vagina presented a mucosa consisting of keratinized squamous epithelium (hyperkeratosis) with an average length of 267.1115 μm . The submucosa was composed of very thick and vascularized connective tissue, and areas containing longitudinal muscular glands that were also vascularized (Fig.2E).

Umbilical cord

This structure presented three vessels (two arteries, one vein) supported by loose mucous connective tissue, or Wharton jelly. These two arteries had a histological structure similar to that of other mammals, but with little definition in the middle layer. All were coated by common endothelium and exhibited an adventitious layer that was continuous with the surrounding tissue (Fig.2F).

Testicles

Manatees 02S0111/42, 02S0111/59, and 01S0111/123 were characterized as newborns owing to their body size (1.19, 1.08, and 1.06m, respectively). Their testes had a structure separated by connective tissue septa, which was richly vascularized with small seminiferous tubules and had sparse cell content, namely spermatogonia and Sertoli

Table 4. Mean \pm SD of mucosal and endometrium thickness in female manatees (*Trichechus manatus*)

Uterus	Mucosal (μm)	Endometrium (μm)
01S0112/12	22.40	1338.34
08S0112/20	19.03	1282.34
01S0112/30	13.91	1221.82
01S0112/31	14.27	1141.44
02S0112/38	18.04	1282.11
02S0112/57	23.86	ND
Mean \pm SD	18.585 \pm 4.083	1253.21 \pm 74.848

ND = no data.

cells. The intralobular connective tissue was rich in spindle cells that sometimes formed concentric blades around the tubules (Fig.3A).

Manatee 02S0111/45 was also classified as a newborn (1.15m), and the general structure of its testis was the same as that of the previously described animals, with a similar general organization of the seminiferous tubules, which were only different in size (Table 5). The intralobular stroma cells did not appear to be predominantly spindle-shaped and had a globular aspect (Fig.3B).

Manatee 02S0111/55 was classified as a juvenile manatee according to the size (2.01m), and the general structure of the testis was also similar to the previous ones. However, there was a clear change in the structure of the lobe, as the seminiferous tubules were similar in size to those of manatee 02S0111/45, but with an evidently increased cell population and presence of spermatogonia, spermatocytes I and II, being characterized as an immature animal. The intralobular stroma had the appearance of mature connective tissue with observable clusters of Leydig cells that were not previously seen in the other animals (Fig.3C).

Manatee 02S0111/73 was considered an adult (3.01m) and the structure of his testis resembled that of other adult mammals. The seminiferous tubules presented a mature and organized production of spermatogonia, spermatocytes I and II, and spermatids with evident spermiogenesis, as confirmed by the presence of spermatozoa in the lumen of the seminiferous tubule. The intralobular stroma was loose and exhibited small clusters of mature Leydig cells (with vacuolized cytoplasm, Fig.3D).

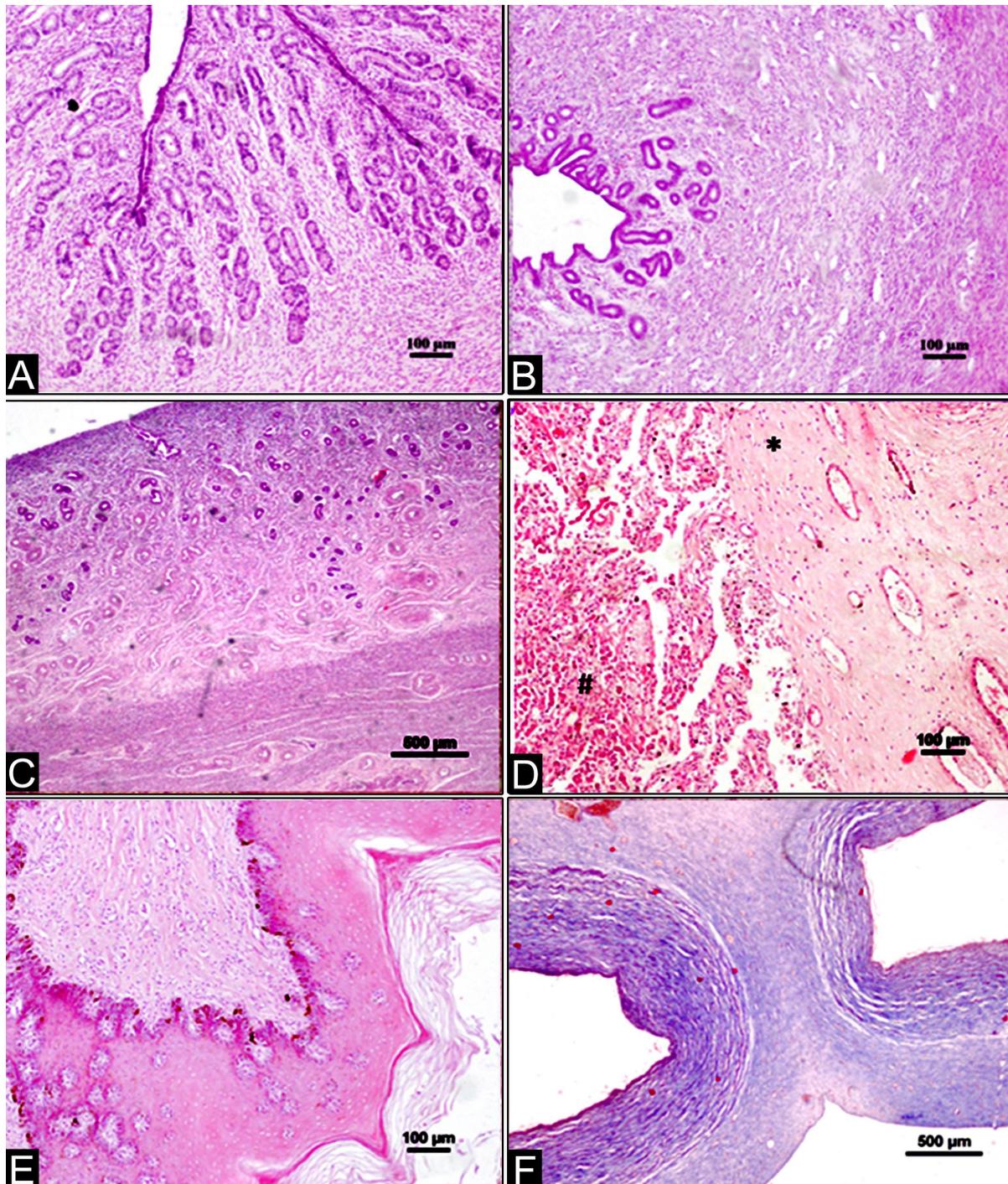


Fig.2. Uteri, placenta, vagina and umbilical cord of *Trichechus manatus*. (A) Uterus in the proliferative phase, with many endometrial glands. HE, obj.40x. (B) Uterus in the secretory phase, with large endometrial glands opening to the light. HE, obj.10x. (C) Uterus in the recovery phase. HE, obj.40x. (D) Placenta with chorionic villi (#) and syncytiotrophoblast (*). HE, obj.10x. (E) Vagina with a keratinized squamous epithelium. HE, obj.10x. (F) Umbilical cord showing the two arteries. Masson's trichrome, obj.40x.

Epididymis

In manatees 02S0111/59 and 02S0111/71, we found multiple tubular structures composed of multiple channels with variable dilation light, coated with pseudo-stratified epithelium and stereocilia, and some portions contained intraepithelial cysts. The wall was composed of abundant fibromuscular tissue, but the stroma contained loose

fibromuscular tissue (Fig.3E). Manatee 02S0111/59 had a less obvious fibromuscular structure.

Penis

Manatees 02S0111/73 and 01S0111/123 presented multiple endothelium-coated vascular channels with variable dilation, filled with blood, in abundant fibromuscular support tissue

Table 5. Mean values of the major diameter (STMa) and minor diameter (STMi) of the seminiferous tubules and mean diameter of the spermatogonia (DS) in male manatees (*Trichechus manatus*)

Testicles	STMa	STMi	DS
02S0111/42	32.692	26.758	3.642
02S0111/45	46.546	39.854	4.363
02S0111/55	56.437	42.041	4.624
02S0111/59	39.778	32.692	4.036
02S0111/73	172.345	134.394	6.162
01S0111/123	35.7680	28.032	ND
Mean ± SD	63.928 ± 53.786	50.629 ± 41.494	4.565 ± 0.965

ND: no data.

with longitudinal, transverse, and oblique fibers. The urethral canal was lined with simple cylindrical epithelium (Fig.3F).

DISCUSSION

This is the first study providing a description of the histological characteristics of the female and male reproductive tracts of manatees (*Trichechus manatus*) from Brazil. Despite the publication of studies on this species in Florida, detailed descriptions of the histological characteristics of the reproductive tract are limited.

We compared the histology of the organs described in this work with that of elephants, which phylogenetically are the

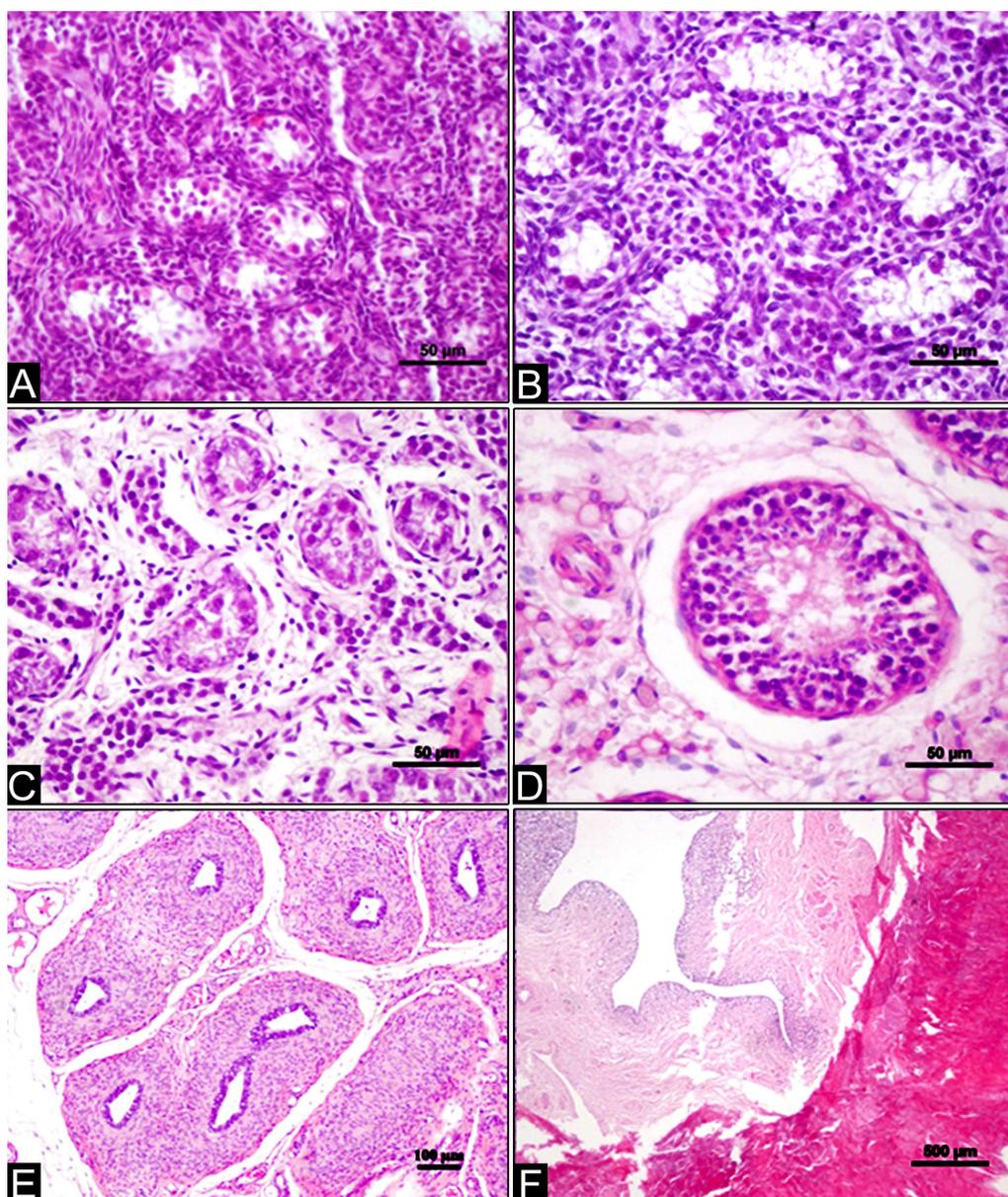


Fig.3. Testis, epididymis and penis of *Trichechus manatus*. (A) Testis showing the spindle cells around the seminiferous tubules. HE, obj.40x. (B) Testis showing the rounded cells around the seminiferous tubules. HE, obj.40x. (C) Testis showing spermatogonia and primary and secondary spermatocytes within the seminiferous tubules. HE, obj.40x. (D) Testis showing complete spermiogenesis within the seminiferous tubules. HE, obj.40x. (E) Epididymis with presence of intraepithelial cysts. HE, obj.10x. (F) Penis in which the cavernous space and the tunica albuginea can be observed. HE, obj.10x.

most closely related group to sirenians, and cetaceans, as a matter of evolutionary convergence.

Female reproductive tract

At the time of necropsy of manatee 02S0112/36, we noticed the absence of ovaries and the presence of tissue in the left antimer, with discrete signs of possible ovarian tissue. Marsh & Kasuya (1984) observed an altered ovary cortex in the short-finned pilot whale (*Globicephala macrorhynchus*), which presented no follicles and had reduced cortex thickness and a large amount of fibrous tissue. This female was lactating and was probably in the puerperal period. The manatee 02S0112/36 also presented a large amount of fibrous tissue in the cortex and almost no follicle, but unlike that of the short-finned pilot whale, the cortex was thicker and, during necropsy, the mammary glands had no increase in volume in the axillary region, or milk.

The uteri of manatees 08S0112/20 and 01S0112/12 were in the proliferative phase, but these animals were neonates, so this characteristic was probably owing to the action of maternal hormones. The uteri of manatees 01S0112/31 and 02S0112/31 were in a proliferative phase, presenting a well-developed epithelium, increased endometrial glands, increased density near the myometrium, and similar features to those of non-pregnant adult female elephants (Hanks & Short 1972).

The uterus of manatee 02S0112/38 was in the recovery phase and belonged to a lactating female, according to the characterization at the time of the necropsy. Histologically, the uterus of this female manatee resembled the uterus of an African elephant female (*Loxodonta africana*) in lactational anestrus, presenting small endometrial glands and low density near the myometrium (Hanks & Short 1972). Coincidentally, one week after the registration of this female, a live newborn was found stranded in the same place. Genetic studies confirmed the relationship between the two animals (Silva 2015).

The placenta was similar to that of Amazonian manatees with a well-developed villus region, syncytiotrophoblast, and the presence of maternal tissue cells and chorionic villi. The umbilical cord was similar to that of *T. inunguis* (Carter et al. 2008).

Male reproductive tract

The general histological appearance of the male reproductive tract in *T. manatus* was similar to that of the African elephant (*L. africana*) and the common dolphin (*Delphinus delphis*). The histological characteristics of the testis were similar to those described for testes of mature and immature animals in Florida manatees (Hernandez et al. 1995).

Murphy et al. (2005) described four stages of testis development in *D. delphis*, and in our work three similar phases were observed. We noticed the immature phase, marked by abundant interstitial tissue and the presence of few spermatogonia; the pubertal stage, in which there is a reduction of the interstitial tissue and a small increase in the diameter of the seminiferous tubules with spermatogonia and spermatocyte; the mature young phase (not observed in this work), which is characterized by the increase of the diameter of the seminiferous tubules and the presence of a central and small lumen; and the sexually mature phase,

marked by the presence of spermatozoa, thus confirming complete spermatogenesis.

According to the classification mentioned above, it was possible to observe that cubs 02S0111/42, 02S0111/45, 02S0111/59, and 01S0111/123 were in the immature phase. The juvenile manatee 02S0111/55 was in the pubertal stage and the adult manatee 02S0111/73 was in the mature phase.

We observed that the mean diameter of the seminiferous tubules varied with the age and size of the manatee, and that smaller and younger animals had smaller, larger tubule diameters, whereas older animals showed larger tubule diameters, similar to what is observed in cetaceans and elephants (Short et al. 1967, Murphy et al. 2005, Westgate & Read 2007).

We also found similarities between the epididymis of the samples studied in this study and the efferent tubules in *L. africana*, which were composed of a ciliary pseudo-stratified epithelium. However, there were no spermatozooids observed in the epididymides, because they belonged to young animals (Short et al. 1967).

CONCLUSIONS

The systematic study of the different components of the reproductive tract, at different stages of growth and sexual development, is necessary to provide a complete profile of the histological characteristics of the reproductive tracts of male and female manatees.

It should be noted that this work appears to be the first to describe the histological characteristics of the tissues of the reproductive tract, placenta, and umbilical cord in this species, which present similar features to those of other aquatic mammals.

The present study seems to contribute significantly to current knowledge of the reproductive histology of the manatee.

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