Mycoplasma agalactiae in semen and milk of goat from Pernambuco State, Brazil¹

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ABSTRACT.- Alves B.H.L.S., Silva J.G., Mota A.R., Campos A.C., Pinheiro Júnior J.W., Santos S.B. & Mota R.A. 2013. *Mycoplasma agalactiae* in semen and milk of goat from Pernambuco State, Brazil. *Pesquisa Veterinária Brasileira 33(11):1309-1312*. Laboratório de Bacterioses dos Animais Domésticos, Departamento de Medicina Veterinária, Universidade Federal Rural de Pernambuco, Av. Dom Manoel de Medeiros s/n, Recife, PE 52171-900, Brazil. E-mail: sanbsantos@gmail.com

In goat and sheep flocks, mycoplasmosis is a disease that may cause severe economical losses associated with polyarthritis, mastitis, agalactia, conjunctivitis, pneumonia and reproductive failure. The latter may involve repeat breeding, granular vulvovaginitis, infertility and abortions. The aim of the present study was to assess the occurrence of Mycoplasma agalactiae (Ma) in semen and milk samples from naturally infected goat in the semiarid region from Pernambuco State, Northeast from Brazil. Thirty-nine semen samples and 81 milk samples were submitted to DNA extraction using a commercially available kit and following the manufacturer's instructions. The polymerase chain reaction (PCR) was then performed in accordance with protocols described in the literature. The results of the present study revealed the presence of Ma in the DNA of 17.9% (7/39) of the semen samples and 3.7% (3/81) of the milk samples. The results obtained in the present study confirm the elimination of the DNA of Ma in the semen and milk samples. The presence of this agent in goat flocks is considered very risky in terms of reproductive disorders and contagious agalactia outbreaks in the Northeast region of Brazil.

INDEXS TERMS: *Mycoplasma agalactiae*, semen, milk, goat, reproductive disorder.

RESUMO.- [*Mycoplasma agalactiae* em sêmen e leite de caprinos do Estado de Pernambuco.] Em caprinos e ovinos as micoplasmoses causam sérias perdas econômicas associadas com poliartrites, mastites, agalaxia, conjuntivite, pneumonias e falhas reprodutivas. Esta última pode envolver repetição de cio, vulvovaginite granular, infertilidade e abortos. O objetivo desse estudo foi verificar a ocorrência de *Mycoplasma agalactiae* (*Ma*) em sêmen e leite de caprinos naturalmente infectados procedentes de regiões semiáridas do Estado de Pernambuco, Nordeste do Brasil.

Foram usadas 39 amostras de sêmen e 81 de leite, as quais foram submetidas à extração do DNA genômico usando um kit comercial, seguindo as instruções do fabricante. A reação da PCR foi realizada de acordo com protocolo previamente descrito na literatura. Os resultados revelaram a presença de DNA de *Ma* nas amostras de sêmen com uma frequência de 17,9% (7/39) e no leite a frequência encontrada foi de 3,7% (3/81). Os resultados obtidos no presente estudo confirmam a eliminação de DNA de *Ma* nas amostras de sêmen e leite analisadas. A presença deste agente nos rebanhos caprinos pode ser considerada um risco para doenças reprodutivas e surtos de agalaxia contagiosa na região Nordeste do Brasil.

TERMOS DE INDEXAÇÃO: *Mycoplasma agalactiae*, sêmen, leite, caprinos, desordens reprodutivas.

INTRODUCTION

Mycoplasmas are bacteria from the *Mollicutes* class and are considered the smallest prokaryotes known without a cell

¹Received on July 19, 2013.

Accepted for publication on September 30, 2013.

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wall (Razin et al. 1998). They are often responsible for severe economical losses in livestock production and in small ruminants. These losses are caused by mycoplasmosis syndromes associated with polyarthritis, mastitis, agalactia, conjunctivitis, pneumonia and reproductive failure, which may involve repeat breeding, granular vulvovaginitis, infertility and abortions (DaMassa et al. 1992). *Mycoplasma agalactiae (Ma)* is the main agent of contagious agalactia (CA). Clinical manifestations occur in the mammary glands, joints and ocular systems and may cause abortions in pregnant goats and sheep (Hasso et al. 1993, Bergonier et al. 1997).

Mycoplasmas cause persistent reproductive failure in the reproductive system of goats and sheep. The species that are most commonly affected are M. bovigenitalium, M. agalactiae, Acholeplasma spp. and Ureaplasma spp. (Kapoor et al. 1984, Nicholas et al. 1999, Nicholas 2002, Gil et al. 2003, Gregory et al. 2012, Santos et al. 2013). The presence of these microorganisms in the reproductive system of males has been associated with diseases such as orchitis, seminal vesiculitis, balanoposthitis, epididymitis and other functional disorders related to the morphology and motility of spermatozoa (Panangala et al. 1981, Pilazec & Truszcynski 1988, Eaglesome et al. 1992, Rizzo et al. 2011). The transmission route is through direct contact during sexual intercourse, artificial insemination or by embryo transfer (Kirkbride 1987, Hasso et al. 1993, Miller et al. 1994). Oliveira (2008) detected a frequency of 72.7% for Mollicutes in sheep semen in São Paulo, Brazil. In Spain, De la Fe et al. (2009) confirmed the excretion of *Ma* in semen samples from goats. The aim of the present study was to detect the DNA of Ma in fresh and frozen semen and milk samples from goats in Brazil.

MATERIALS AND METHODS

Semen and milk samples

A total of thirty-nine semen samples were analyzed, being thirteen of fresh semen and twenty-six of frozen semen. The fresh semen samples were collected from goat bucks of different ages and breeding, healthy or had a history of reproductive disorders, and came from farms in the cities of São José do Egito, Floresta, Sertânia and Afogados da Ingazeira, all of which are in a semiarid region of Pernambuco State, in northeastern Brazil. All of the animals were first submitted to a clinical examination. Subsequently, semen samples were collected using the artificial vagina method. The biological material was tested for macroscopic and microscopic characteristics, in accordance with the guidelines of the Brazilian College of Animal Reproduction (CBRA 1998). Furthermore, twenty-six frozen semen samples were obtained from artificial insemination centers (IAC) in the state of Pernambuco. The frozen semen straws were maintained in liquid nitrogen until usage. Eighty-one milk samples were also collected from goats of different breeding and with lactation stages of different properties. The samples were collected after previous teat flushing with water and soap, drying with paper towels and asepsis with alcohol at 70°GL. A volume of five milliliters of milk was collected in sterile vials (15mL) which were labeled with the name or number of the animal. The fresh semen and milk samples were refrigerated at 4°C in a cooler with ice and transported to the Laboratory of Infectious Diseases - LDIC--DMV/UFRPE.

PCR for Mycoplasma agalactiae

The semen and milk samples were processed for DNA extraction using a commercially available kit, according to the manufacturer's instructions (DNA Easy Blood and Tissues Kit®, Qiagen Biotechnology, guideline page 25). The PCR assays were performed in a mixture prepared with a volume of 25µL, containing 5µL of DNA template, 30pmol of each primer, MgCl₂ (1.5mM), buffer [10mM of Tris-HCl, pH 8.3), a mix of deoxynucleoside triphosphate (50μM), Tag DNA Polymerase (2.5U) and Milli-Q ultrapure water. In-vitro amplifications were performed in the thermo cycler model PTC-100 (MJ-Research®) with primers (MarFor and MarRev) and a thermal profile, based on the protocol described by González et al. (1995), based in variable region V6 of the 16SrRNA gene. The standard, Mycoplasma agalactiae (0.8ng/µL) (Strain BrPB01, Paraiba, Brasil, Gen Bank No JQ612164), was used as a positive control for PCR reactions. Ultrapure water was used as a negative control. The PCR products were analyzed by agarose gel at 1.5% electrophoresis and visualized by staining with Bluegreen® and viewing under ultraviolet light. All analysis was photodocumented.

RESULTS AND DISCUSSION

Prior to semen collection, the goat bucks were clinically examined. None of the animals exhibited clinical symptoms associated with CA infection or lesions in the external genitalia. No abnormalities were found in the analysis of the quality, activity and motility of the semen. Gregory et al. (2012) reported the presence of *Mycoplasma* and *Ureaplas*ma in the semen of sheep after recording no abnormalities in the preliminary clinical examination. Based on the PCR, all fresh semen samples (0/13) were negative for Ma. In goats of the northeast region more investigations are needed for determine the importance of the semen in the transmission of *Ma* during sexual intercourse. In the present study few goat bucks were available and the frequency of Ma may have been underestimated. Therefore, further studies are needed with a larger number of animals to establish the excretion dynamics in the fresh semen of goats. In the microbiological analysis of the fresh semen was possible identify others bacterial agents (unpublished data).

In frozen semen, the frequency of Ma was 26.9% (7/26). This is the first time that Ma in frozen semen of goats has been found in artificial insemination centers in Pernambuco state. Ma in others occasions has been reported in outbreaks of CA in other semiarid regions in Brazil which are currently considered endemic for Ma and CA (Azevedo et al. 2006). Unfortunately, there are no studies of the influence of this microorganism in reproductive diseases.

In Brazil, the significance of mycoplasmosis as a cause of reproductive failure in goats and sheep is unknown. However, functional alterations in the semen of sheep bucks have been reported along with the presence of *Mycoplasma* spp. and *Ureaplasma* spp. (Gregory et al. 2012). Santos et al. (2013) detected a frequency of 26.0% for *Mollicutes* and 12.0% for *Ureaplasma* spp. in the semen of sheep from the Northeast region of Brazil. The role of these asymptomatic goat bucks in the spread of CA in the northeast of Brazil is as yet unknown. However, the transmission of *Ma* through semen has been confirmed in Spain by De la Fe et al. (2009) and in Iraq by Hasso et al. (1993).

The high frequency detected in frozen goat semen may be a risk factor for outbreaks of CA and reproductive disorders in regions of intensive goat and sheep breeding in Brazil. The isolation of *Mollicutes* in treated frozen semen has been reported in bovine (Cardoso 2003, Marques et al. 2011). According Marques et al. (2011) addition of antibiotics to semen before freezing reduce the number of microrganisms, but no eliminate ureaplasmas. Cardoso & Vasconcellos (2004) report that antibiotics currently added to semen diluents are ineffective against *Mollicutes*. These results in frozen semen of goats arouses for veterinarians that frozen semen may be a potential infection route during artificial insemination. Artificial insemination centers must be monitored in a program of mycoplasmosis control in Brazil.

In milk samples, the frequency of Ma DNA, obtained from positive animals, was 3.7% (3/81). The milk frequency detected in the PCR was lower than that found by Azevedo et al. (2006) and by Campos et al. (2009). These authors reported a prevalence of 100.0% and 83.2%, respectively, in small ruminants after an outbreak of CA in Paraiba, Brazil. However, these authors conducted their experiments in properties that had CA outbreaks and a higher frequency of positive animals was expected. Al-Momani et al. (2006) reported a positive Ma frequency of 36% among animals in Jordan, whereas De La Fe et al. (2005) reported a frequency of 40% in Spain. In the present study, the low frequency of Ma in the milk of the goats may indicate different stages mycoplasmosis in the flocks studied, as a chronic process or subclinical only harbor a small number are eliminated in the milk. Moreover, the continuous attempts at treating mastitis and CA with current antibiotics administered for a long period in flocks of the region may have influenced the elimination of agent in the milk in the period of the sampling. In this study was verified clinical and subclinical mastitis in the goat flocks.

CONCLUSION

The results confirmed the elimination of *Ma* DNA in semen samples from goat bucks and in the milk of goats. The present study is the first report of this agent in frozen semen in the Northeast region of Brazil.

Research Ethics Committee.- The present study was approved by the ethics committee of UFRPE/DMV under Proc. 006/2011.

Acknowledgements.- This study was supported by Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco (FACEPE) (Proc. APQ 1512-20 5.05/10 and APQ-1226-5.05/10).

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