

## First identification of natural infection of *Rickettsia rickettsii* in the *Rhipicephalus sanguineus* tick, in the State of Rio de Janeiro<sup>1</sup>

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**ABSTRACT.**- Cunha N.C., Fonseca A.H., Rezende J., Rozental T., Favacho A.R.M., Barreira J.D., Massard C.L., Lemos E.R.S. 2009. First identification of natural infection of *Rickettsia rickettsii* in the *Rhipicephalus sanguineus* tick, in the State of Rio de Janeiro. *Pesquisa Veterinária Brasileira* 29(2):105-108. Curso de Pós-Graduação em Ciências Veterinárias, Universidade Federal Rural do Rio de Janeiro, Seropédica, RJ 23890-000, Brazil. E-mail: [adivaldo@ufrj.br](mailto:adivaldo@ufrj.br)

The Brazilian Spotted Fever (BSF) is a zoonotic disease caused by *Rickettsia rickettsii* and transmitted by ticks of the genus *Amblyomma*, more frequently, *Amblyomma cajennense*. The aim of this paper was to report the first molecular detection of *R. rickettsii* on *R. sanguineus* naturally infected in Rio de Janeiro, Brazil. Ticks were collected from dogs in a rural region of Resende municipality, Rio de Janeiro State, Brazil ( $22^{\circ}30'9.46''S$ ,  $44^{\circ}42'44.29''W$ ), where occurred five human cases of BSF in 2006. The ticks were identified under a stereoscopic microscope and separated in pools by stages, species and sex. DNA extraction was carried out using QIAamp DNA Mini Kit (QIAGEN®). The DNA was submitted to PCR amplification using 04 set of primers: Rr190.70p/Rr190.602n (*OmpA*, 532bp), BG1-21/BG2-20 (*OmpB*, 650bp), Tz15/Tz16 (17 kDa protein-encoding gene, 246bp) and RpCS.877p/RpCS.1258n (*gltA*, 381bp). PCR products were separated by electrophoresis on 1% agarose gels and visualized under ultraviolet light with ethidium bromide. PCR products of the expected sizes were purified by QIAquick® and sequenced by ABI PRISM®. The generated nucleotide sequences were edited with using Bioedit® software and compared with the corresponding homologous sequences available through GenBank, using Discontiguous Mega Blast (<http://www.ncbi.nlm.nih.gov>). It was confirmed *R. rickettsii* by sequencing of the material (GenBank FJ356230). The molecular characterization of *R. rickettsii* in the tick *R. sanguineus* emphasizes the role of dogs as carriers of ticks from the environment to home. Moreover, this result suggests that there is a considerable chance for active participation of *R. sanguineus* as one of tick species in the transmission of *R. rickettsii* to human being in the Brazilian territory.

**INDEX TERMS:** *Rhipicephalus sanguineus*, Acari, Ixodidae, *Rickettsia rickettsii*, molecular analysis, Brazilian spotted fever.

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**RESUMO.-** [Primeira identificação de infecção natural por *Rickettsia rickettsii* no carrapato *Rhipicephalus sanguineus* no Rio de Janeiro.] A Febre Maculosa Brasileira (FMB) é uma zoonose causada por *Rickettsia rickettsii* e transmitida por carrapatos do gênero *Amblyomma*, mais freqüentemente pela espécie *Amblyomma cajennense*. Este trabalho tem como objetivo relatar a primeira detecção molecular de *R. rickettsii* em *Rhipicephalus sanguineus* naturalmente infectado no Rio de Janeiro, Brasil. Carrapatos foram coletados de cães, procedentes de uma região rural do município de Resende, estado do Rio de Janeiro, Brasil (22°30'9.46"S, 44°42'44.29"W), onde ocorreram cinco casos humanos de FMB em 2006. Todos os carrapatos foram identificados segundo chave dicotômica, utilizando-se lupa estereoscópica e separados de acordo com estágio, espécie e sexo. Para a extração de DNA utilizou-se o kit comercial QIAamp DNA (QIAGEN®). O DNA foi submetido à técnica de PCR utilizando 04 conjuntos de iniciadores para a amplificação dos genes: Rr190.70p/Rr190.602n (*OmpA*, 532bp), BG1-21/BG2-20 (*OmpB*, 650bp), Tz15/Tz16 (17 kDa gene que codifica a proteína, 246bp) e RPCs .877p/RpCS.1258n (*gltA*, 381bp). Os produtos da PCR foram separados por eletroforese em gel agarose 1% corados com brometo de etídio e visualizados sob luz ultravioleta e, aqueles que apresentaram bandas amplificadas foram purificados utilizando-se o kit comercial QIAquick® e seqüenciados pelo ABI PRISM®. As seqüências nucleotídicas foram geradas usando Bioedit®, editado em software e comparados os correspondentes homólogos com as sequências disponíveis através GenBank, utilizando Discontiguous Mega Blast (<http://www.ncbi.nlm.nih.gov>). Confirmou-se *R. rickettsii* (GenBank FJ356230) no seqüenciamento de apenas um espécime, adulto de carrapato *R. sanguineus*. A caracterização molecular de *R. rickettsii* em exemplar de carrapato *R. sanguineus* confirma que esta espécie pode ter importante papel na transmissão de *R. rickettsii* para humanos no território brasileiro.

**TERMOS DE INDEXAÇÃO:** *Rhipicephalus sanguineus*, Acari, Ixodidae, *Rickettsia rickettsii*, Análise molecular, Febre maculosa brasileira.

## INTRODUCTION

Brazilian spotted fever (BSF) is a life-threatening rickettsiosis caused by *Rickettsia rickettsii* that is transmitted to human beings by ticks of genus *Amblyomma*, more frequently, *Amblyomma cajennense*, a tick species primarily associated with horses (Dias & Martins, 1939; Oliveira et al. 2000). Although other species of *Rickettsia* have been described in the Brazilian territory, as *Rickettsia parkeri* (Horta et al. 2004, Horta et al. 2007, Labruna et al. 2007) and *Rickettsia felis* (Cardoso et al. 2006, Oliveira et al. 2008), only *R. rickettsii* has been identified in BSF human cases.

Until recently, *Dermacentor andersoni*, *D. variabilis* in North American beside *A. cajennense* in South America were the only species of ticks associated with spotted fever

rickettsioses on the American Continent (Parola et al. 2005). Nowadays, other species have been reported as *A. triste* (Venzal et al. 2008) in Uruguay and *Rhipicephalus sanguineus* in the USA (Demma et al. 2005, Wikswo et al. 2007).

*Rhipicephalus sanguineus*, commonly known as the brown dog tick, is a three-host tick that feeds primarily on dogs and occasionally on other hosts, including human beings (Walker et al. 2000). This tick species is the main vector and reservoir of *Rickettsia conorii* complex that cause Mediterranean spotted fever in Europe, Africa and Asia (Parola et al. 2005).

Although the role of *R. sanguineus* in the transmission cycle of a severe spotted fever rickettsiose to humans have been demonstrated since 1940's in Mexico (Bustamante et al. 1946, Bustamante et al. 1947), only in 2002, *R. rickettsii* was identified in *R. sanguineus* in the USA as important vector of the Rocky Mountain Spotted Fever (RMSF) in Eastern Arizona (Demma et al. 2005)

Burgdorfer et al. in 1975, registered the possibility of *R. sanguineus* involvement in the epidemiology of RMSF, reporting that ticks of this species collected in infected dogs with *Rickettsia* spp. were able to transmit and cause the disease, experimentally.

Recently in Brazil, human parasitism by *R. sanguineus* have been reported (Dantas-Torres et al. 2006, Louly et al. 2006). Till now, no data is published regarding the presence of vector-borne rickettsia in this specie of tick in a BSF endemic area.

On the other hand, recently *R. sanguineus* has been identified as reservoir of *Rickettsia felis* in Brazil (Cardoso et al. 2006, Oliveira et al. 2008), although previous studies had shown evidence of infected *R. sanguineus* in endemic areas in Brazilian states of Minas Gerais and Rio de Janeiro (Lemos et al. 1997a, Rozental et al. 2002)

The increasing number of cases of human parasitism by *R. sanguineus* ticks reported in the literature indicates that the interaction between human beings and *R. sanguineus* may be more common than it is recognized in the American continent.

Usually, *R. sanguineus* parasitizes dogs in urban areas, but sometimes they are found in rural farming areas (Soares et al. 2006, Labruna et al. 2001). In BSF endemic area, the conglomerations of *R. sanguineus* create a great risk to humans whose can be attacked by infected ticks in and around their houses. This study reports the first molecular detection of *R. rickettsii* in *R. sanguineus* ticks in Rio de Janeiro State, Brazil.

## MATERIALS AND METHODS

The material here analyzed is part of an ongoing study that is been developing at Engenheiro Passos District (22°30'165"S, 44°42'654"W), located in the city of Resende, Rio de Janeiro State, Brazil where in May 2006 five BSF cases with two deaths were confirmed by serology. This area is situated in the western portion of the State in Paraíba do Sul middle river valley. The native vegetation, Atlantic Forest, has been replaced by

degraded areas, characterized by the presence of livestock for milk production and current reforestation with *Eucalyptus* spp.

In June 2006, a study was conducted in this district, when ticks were collected from dogs and horses at or near site of confirmed cases of BSF. After identifying through a stereoscopic microscope using taxonomic key (Aragão & Fonseca 1961, Barros-Battesti et al. 2006), the ticks were disinfected by immersion in 70% alcohol followed by washing in sterile water. Subsequently the ticks were separated in pools, by stage, species, sex and source.

DNA was extracted from ticks by freezing in liquid nitrogen and crushing with a sterile micropesle, resuspended with 20 $\mu$ l of sterile brain heart infusion (BHI) and stored at -20°C until nucleic acid extraction. Total DNA was extracted from the pulverized ticks using QIAamp DNA Mini Kit (QIAGEN™). DNA was divided in pools organized by locality, species and sex and were screened for the presence of *Rickettsia* DNA by polymerase chain reaction (PCR) using four set of primers: Rr190.70p/Rr190.602n (*OmpA*) (Regnery et al. 1991), BG1-21/BG2-20 (*OmpB*) (Eremeeva et al. 1994), Tz15/Tz16 (17 kDa protein encoding-gene.) (Tzianabos et al. 1989) and RpCS.877p/RpCS.1258n (*gltA*) (Roux et al. 1997). If a pool demonstrated an expected PCR product, DNA of each tick specimen that made part of that pool was individually tested.

PCR conditions consisted of an initial DNA denaturation and hot start at 95°C for 5min; followed by 40 consecutive cycles of 40 sec denaturation at 95°C, primer annealing at 55°C for 1 min, extension at 72°C for 1min 10sec, and a 7min extension at 72°C. For each reaction, 8  $\mu$ l of the DNA template from each individual tick sample were added to 2.5 $\mu$ l PCR buffer (10X Invitrogen™) buffer, 1.2 $\mu$ l of each primer (20mM), 1.5 $\mu$ l MgCl<sub>2</sub> (3mM), 0.25 $\mu$ l of dNTP mixture (20 mM), 0.25 $\mu$ l Platinum Taq DNA Polymerase (5 U/ $\mu$ l Invitrogen™) and nuclease free water to a final volume of 25 $\mu$ l. 5 $\mu$ l of DNA extracted from *R. rickettsii*-infected *Amblyomma cajennense* ticks was used as positive control. PCR products were stained by ethidium bromide and visualized by electrophoresis in 1% agarose gel.

The PCR products of the expected amplicon size (BG-1-21/BG2-20, *OmpB*, and RpCS.877p/RpCS.1258n, *gltA*) were purified by QIAquick™ and their forward and reverse nucleotide sequences were determined in a DNA sequencer analyzer (ABI Prism 3100 Genetic-Applied Biosystems/Perking Elmer, California, USA). Nucleotide sequence homologies, to determine similarities to other *Rickettsia* species, were established using NCBI BLASTn network service (<http://www.ncbi.nlm.nih.gov/BLAST>).

## RESULTS AND DISCUSSION

From the 1.445 ticks collected, *Rhipicephalus sanguineus*, *Amblyomma cajennense*, *A. aureolatum*, *A. ovale*, *Dermacentor (Anocentor) nitens* and *Rhipicephalus (Boophilus) microplus*, besides immature stages of *Amblyomma* sp. from animals, only 22 were submitted to molecular analysis until now. The result revealed PCR products for one specimen of *R. sanguineus* which showed amplicons to the four set of primers with 99% similarity to *R. rickettsii* (GenBank FJ356230b). The infected tick was identified as an engorged female, collected directly from a male dog, approximately one-year old, of Poodle breed, who lived freely in different environments in the rural property where the BSF cases occurred.

The occurrence of five confirmed cases of BSF in this region and the lack of *A. cajennense* infected by *R. rickettsii*, until now, is in agreement with the data showed by Demma et al. (2005) in which *R. rickettsii* was detected in *R. sanguineus* in an area in Arizona where the main vectors in United States, *D. variabilis* and *D. andersoni* were not identified.

More publications have showed evidence that the brown dog tick is the RMSF vector in some regions of the USA, where close human interaction with free-roaming community dogs heavily parasitized by these ticks is observed. Also, in California, where the role of *R. sanguineus* is unclear as a potential vector of this disease, *R. rickettsii* was detected in this tick, but some differences in genotypes were shown between the strains isolated from Arizona, demonstrating a significant genetic diversity in this species of the spotted fever group rickettsiae SFGR (Wikswo et al. 2007).

Although until now there is not strong evidence that *R. sanguineus* as the vector of BSF, previous studies have shown evidence of involvement of *R. sanguineus* in endemic regions in the states of Minas Gerais and Rio de Janeiro. Hemolymph samples of specimens collected from *R. sanguineus* were analyzed by Giemsa stain and immuno-fluorescence assay and microorganisms with similar morphology to (SFGR) were detected (Lemos et al. 1996, Lemos et al. 1997a, Rozental et al. 2002).

Currently, in Brazil, the main tick implicated in epidemiology of the BSF is *A. cajennense*, an ixodid with low specificity for the vertebrate host, its immature stages commonly are found parasiting human beings in rural areas or at least where primary hosts are found (Lemos et al. 1997b, Oliveira et al. 2000). *A. aureolatum* is another involved tick in the epidemiology of BSF, feeding on wild animals during its immature stages and parasitizing dogs during the adult stage. *R. rickettsii* was identified in this tick, collected from dogs at an endemic region of São Paulo state, suggesting be an important vector in Brazil (Pinter & Labruna 2006).

Dogs from rural areas move freely in pastures, forests and stables around the houses, most of them being pets, mainly for children. Thus, the presence of these animals in homes is a risk factor for the occurrence of human cases, because they carry ticks from their natural environment into the house and/or to the neighboring areas.

Although this result is preliminary, the presence of *R. sanguineus* infected by *R. rickettsii* in a BSF endemic area confirms that this species may play a role in transmitting SFGR to human in Resende, state of the Rio de Janeiro, Brazil, an area where dogs, horses, cattle; domestic cats and their ticks have continuous contact with wild animals such as capybaras, marsupials and small rodents.

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